line  $(10^{-7} \text{ to } 10^{-5} \text{ m})$  inhibited contractions evoked by electrical stimulation (0.3 to 4 Hz) of the postganglionic adrenergic nerves (n = 5) and by exogenous noradrenaline  $(5 \times 10^{-9} \text{ to } 5 \times 10^{-6} \text{ m}, n = 5)$ . Amitriptyline  $(10^{-6} \text{ and } 10^{-5} \text{ m})$  depressed contractions evoked by noradrenaline  $(5 \times 10^{-8} \text{ M})$ to a significantly greater extent (P < 0.05) than contractions of equivalent amplitude evoked by electrical stimulation (0.6 Hz). In superfusion experiments, amitriptyline  $(5 \times 10^{-6} \text{ m})$  increased the overflow of radioactivity and of [3H]-noradrenaline from electrically stimulated (1 Hz) strips of vein both in the absence (n = 5) and presence (n = 6) of cocaine. Thus amitriptyline has a presynaptic effect, unrelated to uptake blockade, which increases the release of noradrenaline from the adrenergic nerve ending. This increase in the stimulated overflow of [3H]-noradrenaline was abolished by phentolamine ( $10^{-5}$  M, n = 6), indicating that it was due to blockade of presynaptic α receptors by amitriptyline.

Exogenous acetylcholine  $(2.7 \times 10^{-7} \text{ M})$  depressed the increase in tension and in the overflow of radioactivity and of [ $^3$ H]-noradrenaline from electrically stimulated (2 Hz) strips of vein (n = 6). This presynaptic inhibitory action of acetylcholine was significantly reduced (P < 0.001) by amitriptyline  $(10^{-6} \text{ M}, n = 6)$ . Exogenous histamine  $(10^{-5} \text{ M})$  also depressed the increase in the overflow of radioactivity from elec-

trically stimulated (2 Hz) veins. This effect was not attenuated by amitriptyline (5  $\times$  10<sup>-6</sup> M, n = 5).

These results demonstrate that amitriptyline in the above concentrations is an antagonist of presynaptic  $\alpha$  and muscarinic receptors. Blockade of these receptors could be important both in the central actions of the drug and in the genesis of the tachycardia and cardiac arrythmias often caused by amitriptyline (Moir, et al., 1972).

#### References

GHOSE, K. & COPPEN, A. (1977). Noradrenaline, depressive illness and the action of amitriptyline. *Psychopharmac.*, 54, 57-60.

Langer, S.Z. (1977). Presynaptic receptors and their role in the regulation of transmitter release. *Br. J. Pharmac.*, **60**, 481-497.

McGrath, M.A. (1977), 5-hydroxytryptamine and neurotransmitter release in canine blood vessels. *Circulation Res.*, 41, 428-435.

MOIR, D.C., CROOKS, J., CORNWELL, W.B., O'MALLEY, K., DINGWALL-FORDYCE, I., TURNBULL, M.J. AND WEIR, R.D. (1972). Cardiotoxicity of amitriptyline. *Lancet*, 2, 561-564.

SCHILDKRAUT, J.J. (1965). The catecholamine hypothesis of affective disorders: a review of supporting evidence. Am. J. Psychiat., 122, 509-522.

# Preliminary characterisation of the presynaptic receptor for 5-hydroxytryptamine in dog isolated saphenous vein

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We have shown that 5-hydroxytryptamine (5-HT) inhibits contractile responses induced by electrical stimulation in dog saphenous vein and have suggested that this effect is mediated via a specific pre-synaptic 5-HT receptor (Feniuk, Humphrey & Watts, 1978). This receptor like the 5-HT receptor situated post-synaptically in this preparation, is only weakly blocked by the 5-HT antagonist methysergide (Feniuk, Humphrey & Watts, 1979). Indeed methysergide appears to be a partial agonist at the postsynaptic receptor (Apperley, Humphrey & Levy, 1977) and we have therefore investigated whether methysergide has agonistic activity on the presynaptic receptor.

Saphenous veins were removed from barbitone

anaesthetised dogs and cut spirally into strips. These were stimulated electrically in vitro as described previously (Feniuk, et al., 1978, 1979) and the inhibitory effects of 5-HT and methysergide examined. In other experiments we studied electrically stimulated tritium release following incubation with 1-[7-3H]-noradrenaline (10 m Ci/0.67  $\times$  10<sup>-6</sup> mol/l) at 37°C for 2 h in a modified Krebs solution (Apperley, Humphrey & Levy, 1976) containing ascorbic acid (1.1  $\times$  10<sup>-4</sup> mol/l) and disodium EDTA (4.0  $\times$  10<sup>-6</sup> mol/l). Each strip was then transferred to a bath and washed to remove extracellular tritium using Krebs solution containing ascorbic acid, disodium EDTA, cocaine  $(3.0 \times 10^{-5} \text{ mol/l})$ , indomethacin  $(2.8 \times 10^{-6} \text{ mol/l})$ and corticosterone  $(4.0 \times 10^{-5} \text{ mol/l})$ . Subsequently the bath was drained and refilled every 2 min and the amount of radioactivity in the Krebs measured by liquid scintillation counting. Each strip was stimulated electrically for 2 min at 2 Hz every 20 minutes. In some experiments [3H]-noradrenaline was separated from the other tritiated metabolites using paper chromatography (Levin, 1973).

5-HT and methysergide inhibited contractions induced by electrical stimulation producing 50% inhibition at  $3.2 \pm 0.6 \times 10^{-8}$  mol/l (n = 20) and  $1.7 \pm 0.4 \times 10^{-6}$  mol/l (n = 24) respectively (mean values  $\pm$  s.e. mean) The inhibitory effects of 5-HT and methysergide were potentiated by cyproheptadine ( $1.0 \times 10^{-6}$  mol/l) but were unaffected by haloperidol ( $1.0 \times 10^{-6}$  mol/l), propranolol ( $1.0 \times 10^{-6}$  mol/l), atropine ( $1.0 \times 10^{-6}$  mol/l), mepyramine ( $1.0 \times 10^{-6}$  mol/l) or cimetidine ( $1.0 \times 10^{-5}$  mol/l). 5-HT ( $1.0 \times 10^{-7}$  mol/l) and methysergide ( $3.0 \times 10^{-6}$  mol/l) inhibited the electrically stimulated release of tritium by  $78 \pm 4\%$  (n = 6) and  $47 \pm 7\%$  (n = 6) respectively and these inhibitory effects were not antagonised by phentolamine ( $1.0 \times 10^{-6}$  mol/l) which itself increased tritium release by about four fold, 5-HT and methysergide also inhibited the electrically stimulated release of  $[^3H]$ -noradrenaline.

Our results show that methysergide inhibits release of noradrenaline from noradrenergic nerves in dog saphenous vein possibly by activating the presynaptic receptor for 5-HT. This suggests that the presynaptic 5-HT receptor is similar to the postsynaptic 5-HT receptor in this preparation and that both are differ-

ent from the classical D-receptor (see Apperley, Humphrey & Levy, 1977).

#### References

APPERLEY, EIRA, HUMPHREY, P.P.A. & LEVY, G.P. (1976). Receptors for 5-hydroxytryptamine and noradrenaline in rabbit isolated ear artery and aorta. *Br. J. Pharmac.*, 58, 211-221.

APPERLEY, EIRA, HUMPHREY, P.P.A. & LEVY, G.P. (1977). Two types of excitatory receptor for 5-hydroxytryptamine in dog vasculature? Br. J. Pharmac., 61, 465P

FENIUK, W., HUMPHREY, P.P.A., & WATTS, A.D. (1978). Evidence for a presynaptic inhibitory receptor for 5-hydroxytryptamine in dog isolated saphenous vein. *Br. J. Pharmac.*, 63, 344P-345P.

FENIUK, W., HUMPHREY P.P.A., & WATTS A.D. (1979). Presynaptic inhibitory action of 5-hydroxytryptamine in dog isolated saphenous vein. Br. J. Pharmac. (In press.) Levin, J.A. (1973). Paper chromatographic assay of [3H]-norepinephrine and its five major metabolites. Anal. Biochem., 51, 42-60.

## Evidence for an autoreceptor-mediated presynaptic control of serotonin release in central nerve endings

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The release of noradrenaline from peripheral and central nerve endings is modulated by presynaptic α-autoreceptors (Langer, 1977; Starke, 1977). We have investigated whether a similar mechanism existed in serotonergic terminals, by analyzing the effect of extra-cellular 5-hydroxytryptamine (5-HT) on the release of [<sup>3</sup>H]-5-HT previously accumulated by hypothalamic synaptosomes.

Crude synaptosomes, prepared from adult male Wistar rats, were prelabelled with [³H]-5-HT (0.1 µM, 10 min at 37°C) and distributed in parallel superfusion chambers (Raiteri, Angelini & Levi, 1974). The superfusion media contained chlorimipramine (Cl-IMI, 5 µM) to prevent 5-HT uptake. 5-HT was added after 10 min superfusion with Cl-IMI-containing Krebs-Ringer medium and, 8 min later, the synaptosomes were depolarized with KCl (15 mM). Serotonin antagonists were present from the beginning of superfusion. The superfusion rate was 0.5 ml/min. The [³H]-5-HT in each 1-min fraction and in the synaptosomes at the end of superfusion was measured after isolation on Biorex columns.

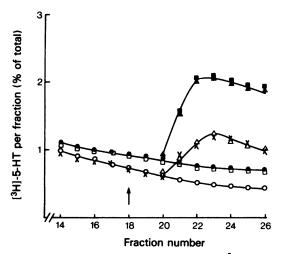


Figure 1 Spontaneous and high K<sup>+</sup>-induced release of [³H]-5HT from hypothalamic synaptosomes: inhibition by extracellular 5HT and antagonism between 5HT and methiothepin. (•) control (Cl-IMI present); (Ο) 0.5 μM 5HT; (□) 0.5 μM 5HT + 0.5 μM methiothepin; (Δ) 15 mM KCl; (Δ) 15 mM KCl + 0.5 μM sHT; (■) 15 mM KCl + 0.5 μM 5HT + 1 μM cyproheptadine. Methysergide (1 μM) and mianserin (1 μM) behaved as cyproheptadine and are not shown in the Figure. The [³H]-5HT present in each fraction is expressed as a percentage of the total [³H]-5HT recovered (fractions + synaptosomes). The curves presented are averages of 3-5 quadruplicate experiments.